



OPENING OF THE PHORBIN EXOCYCLE BY WEAK O-NUCLEOPHILES. STUDY AND OPTIMISATION OF FREE CHLORINE e_6 SYNTHESIS METHODS

D. R. Karimov

Dmitry R. Karimov, Candidate of Chemical Sciences, Associate Professor

Ivanovo State University of Chemistry and Technology, Ivanovo, Russia, dmitriy.karimov@list.ru

Keywords:

phorbines, O-nucleophiles;
exocycle uncoupling,
chlorine e_6 derivatives,
photosensitisers

Abstract. The article considers issues concerning the reaction of exocycle opening in phorbin molecules (on the example of methylpheophorbide a) under the O-nucleophiles - water and alcohols action. According to the author, under certain conditions relatively weak nucleophiles as water and alcohols can cause the opening of the exocycle in the molecule of methylpheophorbide a with the formation of chlorine derivatives e_6 . The study of this reaction, in addition, reveals new synthetic methods to obtain chlorine e_6 esters of different degrees of substitution, and eventually - free chlorine e_6 in the form of triacids. Free chlorine is a valuable photosensitiser for photodynamic therapy (PDT), and a starting compound for the preparation of other photosensitizers. The article also discusses possible synthetic approaches to obtain free chlorine e_6 .

For citation:

Karimov, D.R. (2023) Opening of forbinexocycle by weak O-nucleophiles. Study and optimisation of free chlorine e_6 synthesis methods, *From Chemistry Towards Technology Step-By-Step*, 4(4), pp. 145-157 [online]. Available at: <http://chemintech.ru/index.php/tor/2023-4-4>

Introduction

Some medical issues remain topical and unresolved even in the 21st century. These are, in particular, the issues of cancer and antibiotic-resistant bacterial infections [1-8]. This fact predetermines the constant search for new effective and at the same time maximally safe ways to solve these problems [7]. Photodynamic therapy (PDT) is one of the most promising methods aimed at combating both cancer diseases and pathogenic bacterial microflora; a method associated with the use of photosensitisers (PS) [7-10]. A photosensitizer is usually an organic dye with the ability to selectively accumulate in tumour tissues or bacterial cells. Moreover, during irradiation with light of a certain wavelength, it generates active (singlet) oxygen. Active oxygen, in turn, destroys surrounding tumour tissues or bacterial cell organelles [7]. Since singlet oxygen is generated only in the place of photosensitizer accumulation during



its laser irradiation. The healthy tissues around the tumour or the focus of bacterial infection are not affected [7]. However, photosensitisers have a number of requirements (simultaneous water- and fat-solubility (amphiphilicity), ability to absorb light quanta in the red and near-infrared regions of the spectrum, efficient generation of singlet oxygen and other reactive oxygen species, tropism to tumour cells, low toxicity and phototoxicity, high rate of elimination from the body), etc. Indeed, very few of them can be realised through one chemical compound only. Thus, there is no "ideal photosensitizer" [7]. Therefore, scientist are in the constant search for new photosensitisers - natural and synthetic ones.

Semisynthetic derivatives of natural chlorophyll - derivatives of phorbins, as well as amides and esters of chlorine e_6 (both in free form and as conjugates with various monomeric biomolecules and biopolymers) are very promising as photosensitisers [11-16]. Nevertheless, even these compounds still do not fully satisfy the whole set of requirements for photosensitisers. Therefore, the search for new ways of their synthesis and modification continues. It is also a challenge to obtain free chlorine e_6 in pure form by standard methods, such as alkaline hydrolysis of its esters or methylpheophorbide *a*. It produces a mixture of similar compounds which must be separated [17].

The initial compound for the preparation of chlorine e_6 various derivatives is methylpheophorbide *a*. It is obtained from natural chlorophyll *a* in two steps. The first step is demetallisation (removal of the magnesium ion by hydrochloric acid), and the second one is transesterification (replacement of the phytol residue with a methyl group at position 17 by methanol-sulphuric acid (5% vol.)). In order to obtain chlorine e_6 derivatives from methylpheophorbide e_a , it is necessary to open the phorbinoxocycle. The exocycle opening reaction is a nucleophilic substitution at the carbonyl carbon atom at position 13(1) [18]. The tendency of the exocycle to open under the action of nucleophilic reagents is primarily due to the removal of steric stresses (distortion of valence angles in the exocycle and repulsion of closely located bulk substituents at positions 17 and 13(2)) during the reaction. It also concerns with the peculiarities of the process mechanism (the possibility of negative charge delocalisation in the carboanion formed by heterolytic bond breaking 13(1)-C-13(2)-C) [16].

The reaction of exocycle opening has been studied quite well on the example of the methylpheophorbide *a* interaction with amines acting as N-nucleophiles [18, 19]. The spatial structure of amine (the degree of the nitrogen atom shielding by substituents) and the electronic structure of the macrocycle (the presence of electron-acceptor substituents should hypothetically facilitate the reaction) have a significant influence on reaction proceeding [18]. In this case, the steric factor plays a dominant role and cannot be compensated by the electronic one. Hence, primary, and secondary aliphatic amines react smoothly. In case of secondary amines, the substituents at the nitrogen atom should not be more voluminous than the *n*-propyl group [18]. Moreover, the process characterized by the excesses of amines (100-200-fold). At the same time, there are studies [20, 21] describing the opening of the exocycle by bulk amine molecules with minimum excesses. However, the use of more stringent



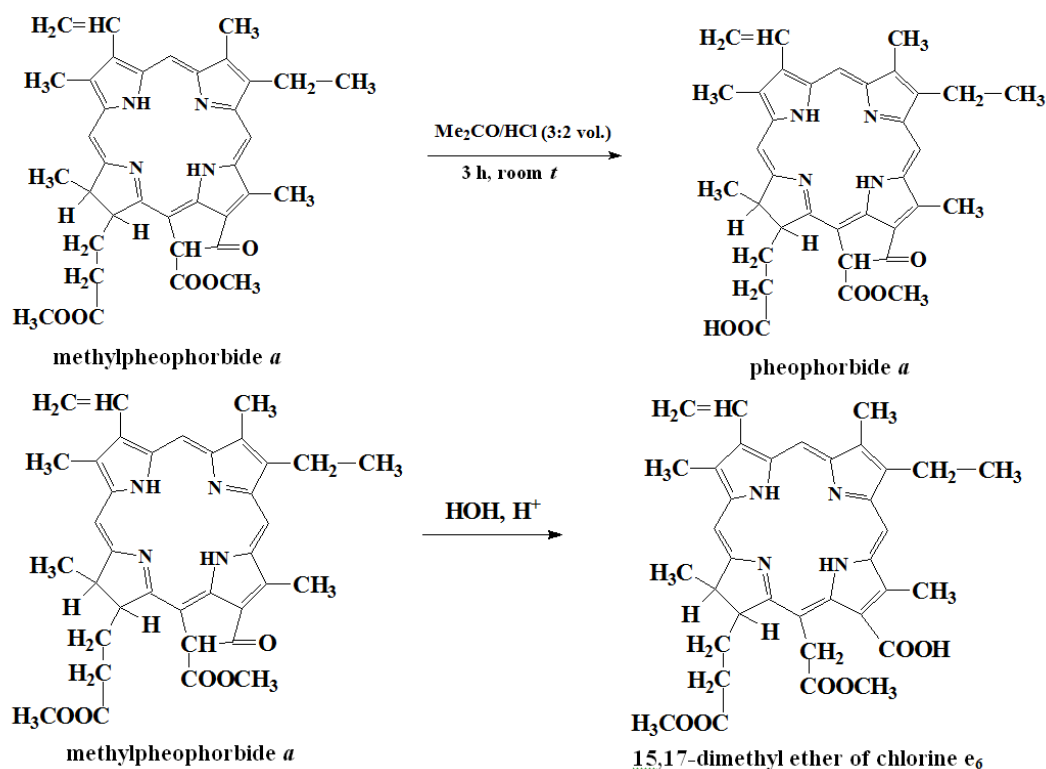
conditions for the reaction of exocycle opening (boiling in toluene instead of room temperature) causes reaction direction changing. Furthermore, amidation of the exocycle ester group occurs without its opening [18].

The opening of the exocycle by O-nucleophiles in methylpheophorbide *a* and its derivatives has been described in the literature [22, 23]. However, those O-nucleophiles should be by very strong ones (in particular, sodium methylate). Alkaline hydrolysis of phorbin derivatives is preparatively inexpedient. Therefore, it causes the formation of a difficult-to-separate mixture of free chlorine e_6 and its methyl esters with various degrees of substitution [18]. For preparation of trimethyl ether of chlorine e_6 , treatment of such mixtures with diazomethane is usually used [22, 23]. Their preparation is associated with certain difficulties (relatively inaccessible reagents) and risks (starting substances for the synthesis, particularly N-nitrosomethylurea, are carcinogenic, and diazomethane itself is poisonous and explosive). The trimethyl ether of chlorine e_6 can be obtained by treatment of methylpheophorbide *a* with sodium methylate [22]. It is also formed by the action of potassium hydroxide solution in methanol on methylpheophorbide *a* in pyridine [24] or tetrahydrofuran. Water, although in trace quantities, was found to be a weak nucleophile, when interacting with phorbins under certain conditions, can form exocycle opening products. Pheophorbide *a* also undergoes exocycle opening upon treatment with potassium hydroxide in the THF-methanol system at room temperature to form 17-carboxy-13,15-dimethyl ether of chlorine e_6 .

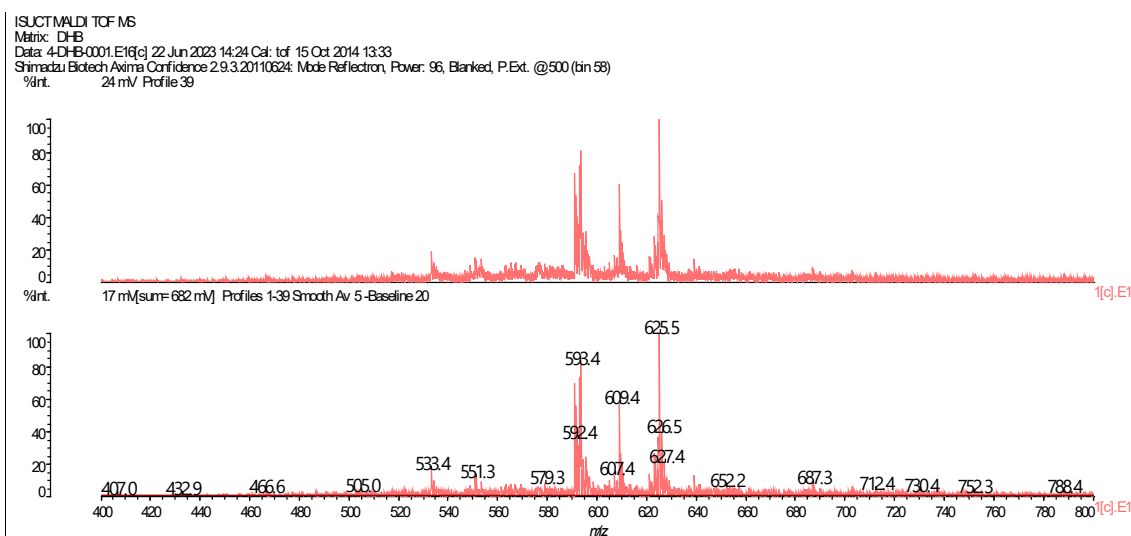
Free chlorine e_6 is formed along with other products when a solution of methylpheophorbide *a* in THF is treated with aqueous potassium hydroxide solution at boiling. One of the by-products is chlorine e_4 . It is a product of a single decarboxylation of chlorine e_6 at position 15 of the macrocycle. The decarboxylation reaction of chlorine e_6 is described in the literature [25]. It proceeds upon heating of chlorine e_6 in basic media.

Experimental part

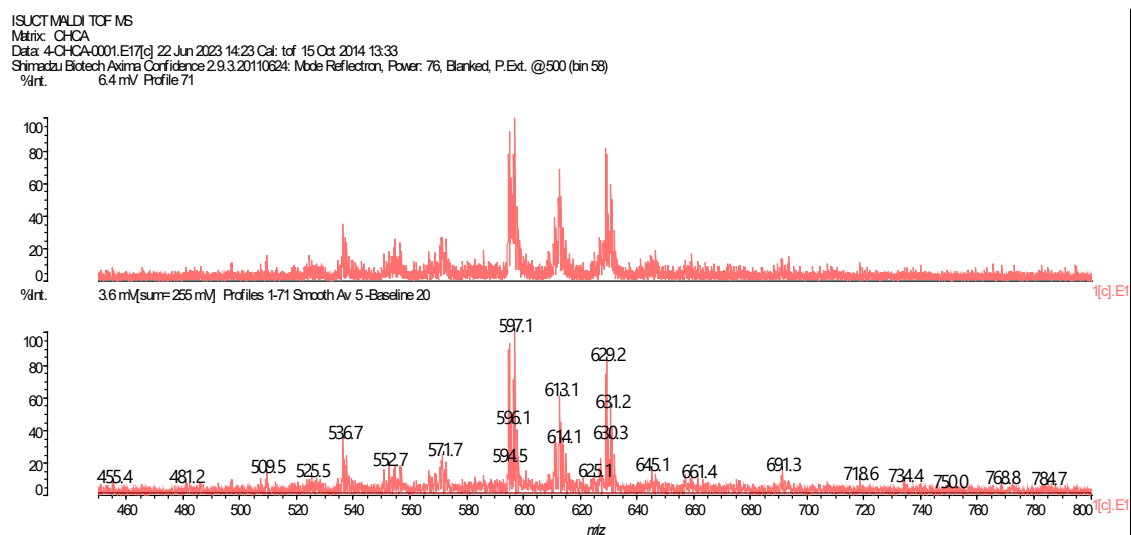
Methylpheophorbide *a* was prepared according to [26]. The solvents were purified by generally accepted methods. Methanol was dried by distillation over magnesium metal; tetrahydrofuran and chloroform were distilled after soaking over potassium hydroxide and sodium sulphate, respectively. Acetone was subjected to distillation with a deflagmator. Potassium hydroxide and hydrochloric acid were used without prior preparation. Mass spectra were recorded using an Axima Confidence time-of-flight tandem mass spectrometer with matrix-assisted ionisation (MALDI TOF) (Shimadzu, Japan). 2,5-dihydroxybenzoic acid (DHB) and α -cyano-4-hydroxycinnamic acid (CHCA) were used as matrix. ^1H NMR spectra were recorded on an Avance-500 spectrometer (Bruker, Germany) with an operating frequency of 500 MHz at 293 K.

1. Study of methylpheophorbide *a* hydrolysis in an acidic medium.

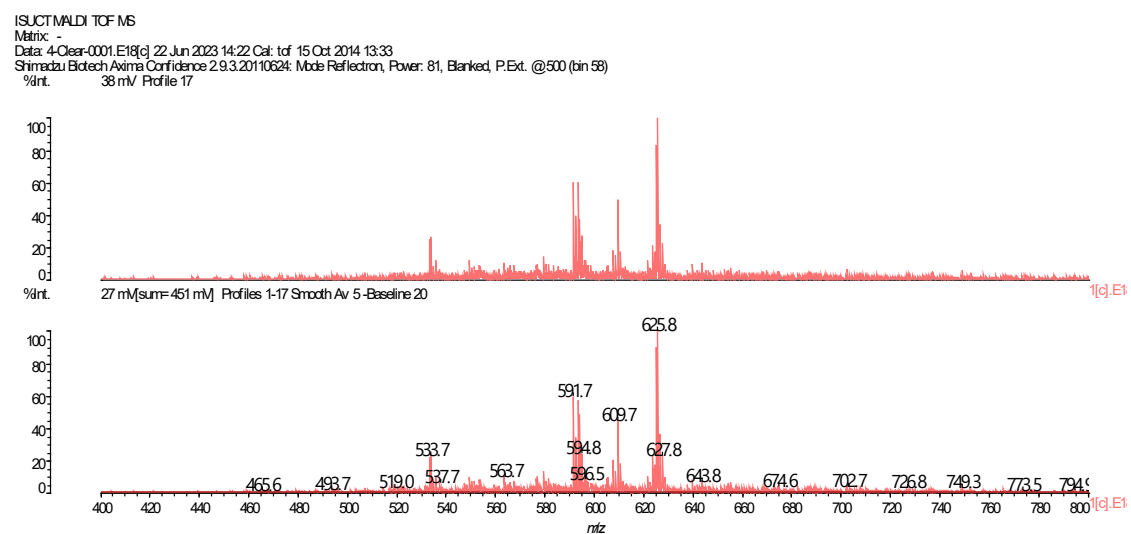
We dissolved methylpheophorbide *a* (200 mg, 0.329 mmol) in 6 ml of acetone. We added 4 ml of concentrated hydrochloric acid to the resulting solution. We stirred the reaction mixture at room temperature for 4 h, followed by extraction with chloroform and methanol, and washing the organic layer with water. TLC (silica gel, dichloromethane - methanol (10% vol.)) of the reaction mixture after extraction shows, in addition to the main zones (pheophorbide *a* as the main product and traces of unreacted initial methylpheophorbide *a*), also the presence of a green-coloured hard-to-suspend fraction. We performed column chromatography of the obtained product in silica gel using chloroform-methanol mixtures (2-20% vol.) as eluent. The green-coloured fraction released the column last and was washed off with chloroform - methanol (20% vol.) mixture. According to mass spectrometry data, this fraction is a mixture of free chlorine *e*₆ and its methyl esters of different degrees of substitution. However, the predominant component of the mixture is dimethyl ester of chlorine *e*₆ ($M = 625$) (Fig. 1). Logically, it is 15,17-dimethyl ether of chlorine *e*₆ with a free carboxyl group at position 13, at the exocycle opening site; the exocycle opening agent is water. The mixture also contains the monomethyl ester of chlorine *e*₆ ($M = 611$) (see Fig. 1), apparently being a 15-monomethyl ester, since the 17-COOCH₃-group undergoes acid hydrolysis more readily than the 15-COOCH₃-group. This follows from numerous experimental data on the preparation of 17-monocarboxylic derivatives of chlorine *e*₆, 13-amides of 15-monomethyl ethers, as well as pheophorbide obtained by selective hydrolysis of the 17-COOCH₃-group in the methylpheophorbide molecule to the 17-COOH-group while retaining the 15-COOCH₃-fragment. Finally, free chlorine *e*₆ corresponds to the 597 signal present in the mass spectra (see Fig. 1).



DHB

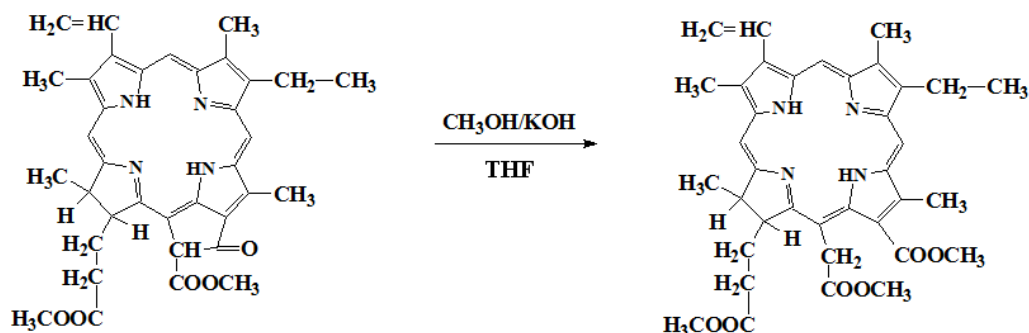


CHCA



Without matrix

Fig. 1. Mass spectra of the "green fraction" on and without different matrices

2. Synthesis of 13,15,17-trimethyl ether e_6 chlorine.

We dissolved methylpheophorbide *a* (121.6 mg, 0.2 mmol) in 6 ml of tetrahydrofuran. We dissolved potassium hydroxide (336 mg, 6 mmol) in 12 ml of methanol. We added the potassium hydroxide solution to the solution of methylpheophorbide *a*; the reaction mixture turned green. We added chloroform and water to the reaction mixture; there was a separation into two layers. The lower coloured organic layer was separated and washed off with water. Column chromatography of the crude product obtained was performed in silica gel using chloroform-methanol mixture (1% vol.) as eluent. The first fraction (green colour) represents the target product. It is confirmed by mass spectrometry ($M = 640$) (Fig. 2) and $^1\text{H-NMR}$ data.

13,15,17-Trimethyl ether of chlorine e_6 . Dark green powder. Yield 89.6 mg (70%). $^1\text{H-NMR}$ (500 MHz, CDCl_3) δ , ppm: 9.71 (s, 1H, 10-H), 9.58 (s, 1H, 5-H), 8.76 (s, 1H, 20-H), 8.08 (dd, 1H, 3(1)-H), 6.37 (dd, 1H, 3(2)-H-*trans*), 6.16 (dd, 1H, 3(2)-H-*cis*), 5.37 (d, 1H, 15(1)-CH), 5.26 (d, 1H, 15(1)-CH'), 4.47 (dd, 1H, 18-H), 4.42 (d, 1H, 17-H), 4.28 (s, 3H, 13-COOCH₃), 3.84 – 3.76 (overlapping m + s, sums. 5H, 8(1)-CH₂, 15(3)-CH₃), 3.66 (s, 3H, 12(1)-CH₃), 3.60 (s, 3H, 17(4)-CH₃), 3.49 (s, 3H, 2(1)-CH₃), 3.32 (s, 3H, 7(1)-CH₃), 2.58 (m, 1H), and 2.22 (m, 2H): 17(1)-CH₂, 17(2)-CH₂; 1.80–1.71 (overlapping d + t + m, sums. 7H, 8(2)-CH₃, 18(1)-CH₃, 17-CH₂), -1.28 (s, 1H, 21-NH), -1.44 (s, 1H, 23-NH).

ISUCTMALDI TOF MS
Matrix: CHB
Data: I-CHB-0001_A19[e] 14 Jul 2023 12:31 Cal: tcf 15 Oct 2014 13:33
Shimadzu Biotech Axims Confidence 2.9.3.20110824: Mode Reflectron, Power: 67, Blanked, P.Ext. @600 (tin 58)
%Int. 86 mV Profile 21

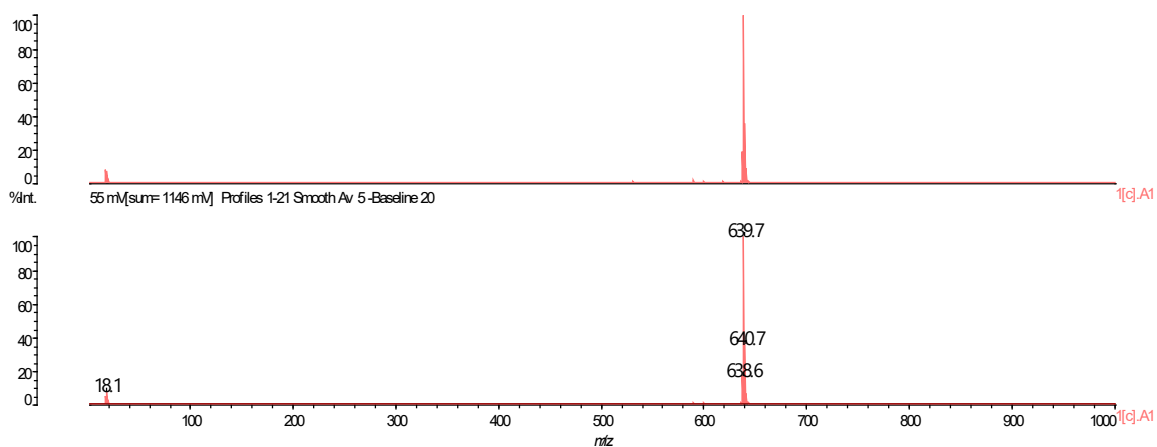


Fig. 2. Mass-spectrum of 13,15,17-trimethyl ether e_6 chlorine

According to mass spectrometry, mono- and dimethyl esters of chlorine e_6 are also formed in trace amounts as by-products ($M = 612$ and 625 , respectively) (Fig. 3). They can be



easily separated chromatographically (washed off the column only by high concentrations of methanol up to 100%).

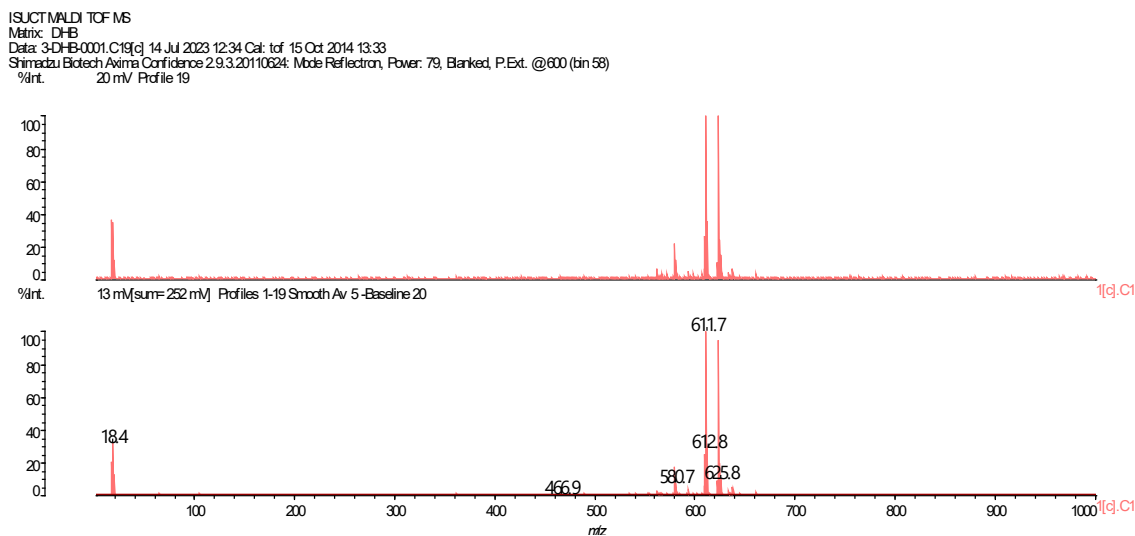
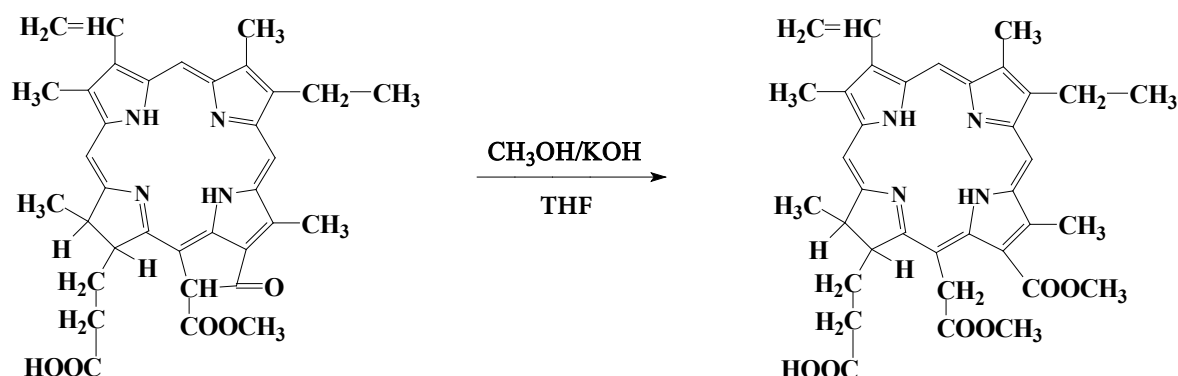


Fig. 3. Mass spectrum of a mono- and dimethyl ester mixture of chlorine e₆

3. Synthesis of 17-carboxy-13,15-dimethyl chlorine ester e₆.



We dissolved pheophorbide *a* (59.4 mg, 0.1 mmol) in 4 ml of tetrahydrofuran and potassium hydroxide (145.9 mg, 2.6 mmol) in 6 ml of methanol. We added the alkali solution to the pheophorbide solution. We diluted the resulting solution (green colour) with water and neutralised it with a calculated amount of dilute hydrochloric acid (0.2 ml of HCl in 20 ml of water). It resulted by coloured precipitation, which was filtered on a paper filter, dried at room temperature, and subjected to column chromatography in silica gel using chloroform-methanol mixture (2% vol.) as eluent. According to mass spectrometry data and ¹H-NMR (M = 626.1 (Fig. 4), first green-coloured fraction represents the target product.

17-Carboxy-13,15-dimethyl ether of chlorine e₆. Dark green powder. Yield 21.1 mg (34%). ¹H-NMR (500 MHz, CDCl₃) δ, ppm: 9.71 (s, 1H, 10-H), 9.58 (s, 1H, 5-H), 8.77 (s, 1H, 20-H), 8.08 (dd, 1H, 3(1)-H), 6.37 (dd, 1H, 3(2)-H-*trans*), 6.16 (dd, 1H, 3(2)-H-*cis*), 5.34 (d, 1H, 15(1)-CH), 5.28 (d, 1H, 15(1)-CH'), 4.46 (overlapping d + dd, sums. 2H, 18-H, 17-H), 4.26 (s, 3H, 13-COOCH₃), 3.80 (qu, 2H, 8(1)-CH₂), 3.76 (s, 3H, 15(3)-CH₃), 3.59 (s, 3H, 12(1)-CH₃), 3.49 (s, 3H, 2(1)-CH₃), 3.31 (s, 3H, 7(1)-CH₃), 2.62, and 2.22 (both – m, sums. 4H, 17(1)-CH₂, 17(2)-CH₂), 1.77 (d, 3H, 8(2)-CH₃), 1.73 (t, 3H, 18(1)-CH₃), -1.45 (s, 1H, 23-NH).



As a by-product, monomethyl ether of chlorine e_6 ($M = 612$) is also formed. It was washed off from the column (together with residual amounts of the target product) by chloroform-methanol mixture (1:1 vol.), as can be seen from mass spectrometry data (Fig. 5).

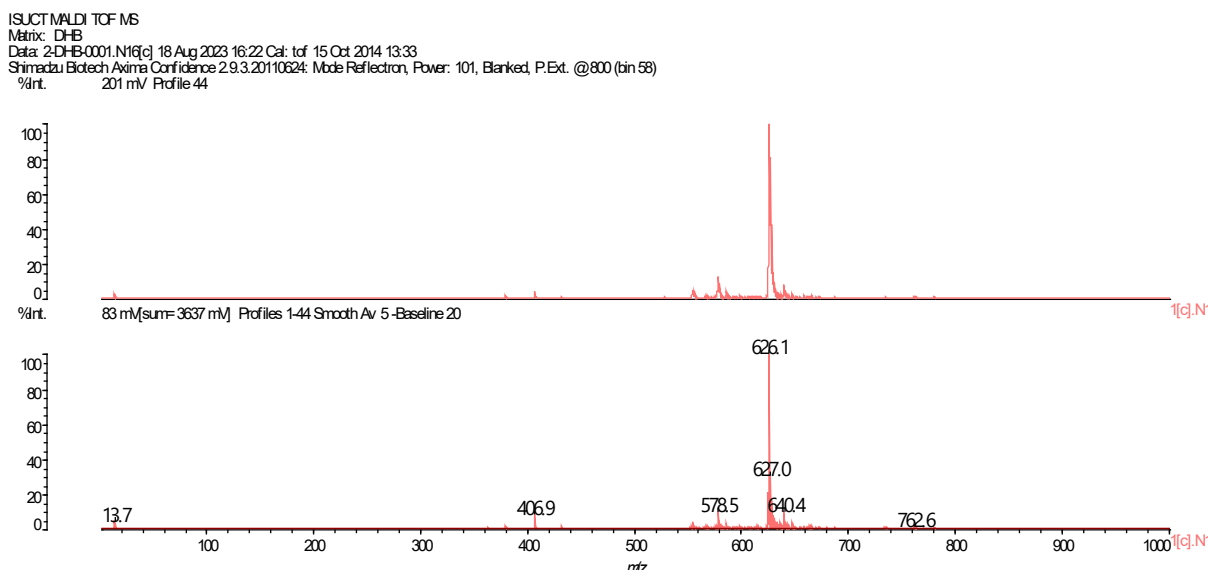


Fig. 4. Mass-spectrum of 17-carboxy-13,15-dimethyl chlorine ester e_6

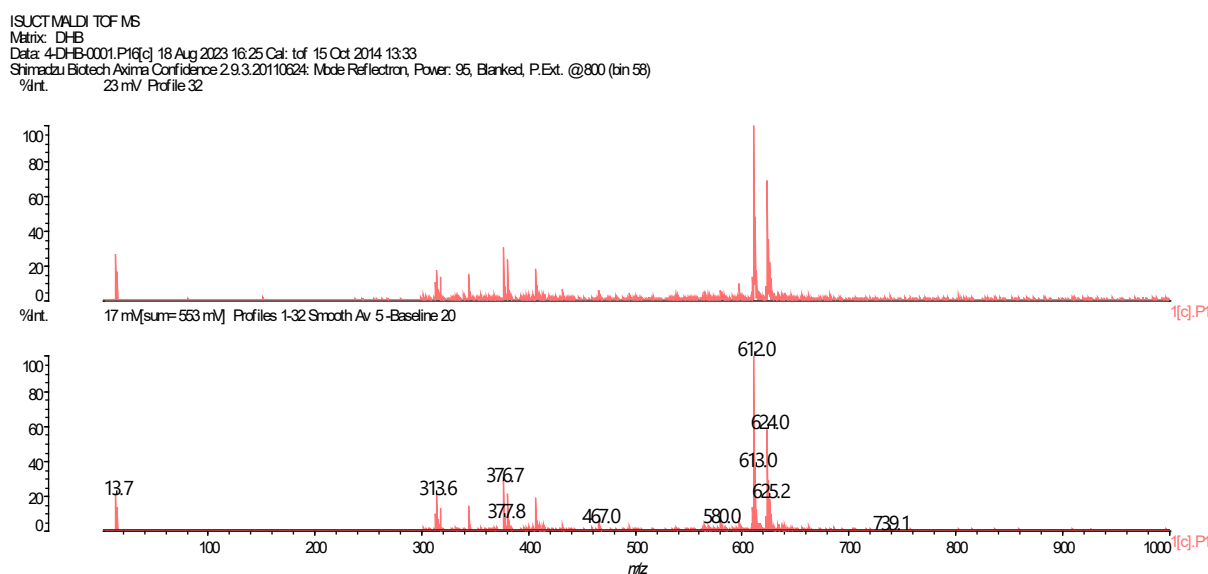
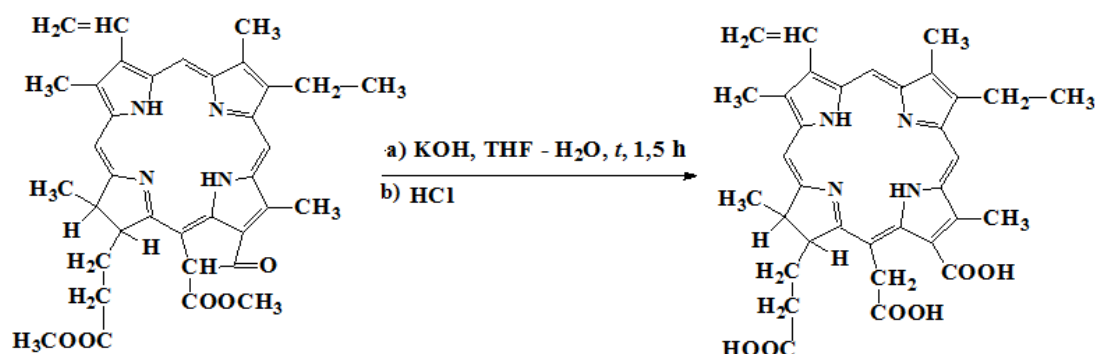


Fig. 5. Mass spectrum of the side fraction in the synthesis of 17-carboxy-13,15-dimethyl ether of chlorine e_6

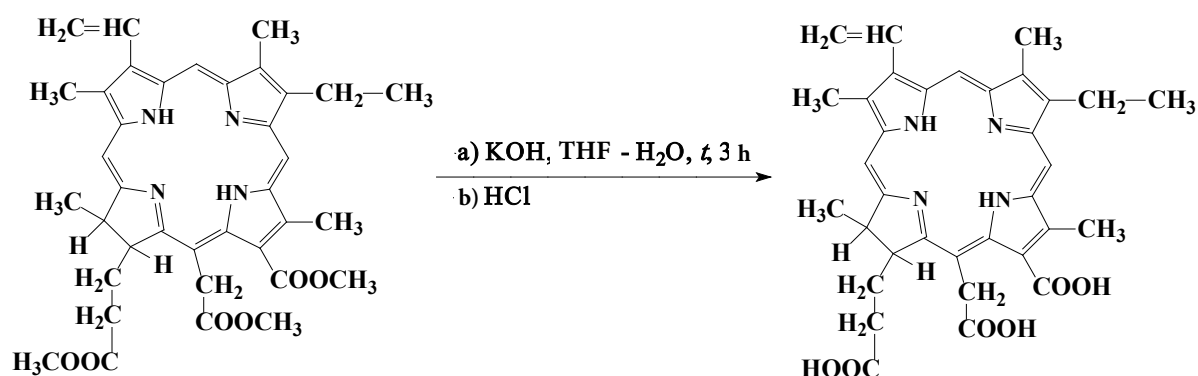
4. Synthesis of chlorine e_6 from methylpheophorbide *a*.





Methylpheophorbide *a* (121.6 mg, 0.2 mmol) was dissolved in 10 mL of tetrahydrofuran; potassium hydroxide (3.36 g, 60 mmol) was dissolved in 10 ml of water. We added the alkali solution to the macrocyclic compound. We boiled the reaction mixture with a reflux condenser for 1.5 h, diluted it with water, and neutralised it with dilute hydrochloric acid (5 ml HCl_{conc.} in 50 ml water). The precipitate was filtered and dried at room temperature. Column chromatography of the obtained product was performed in silica gel using chloroform-methanol mixtures as eluent. Chlorine *e*₆ elutes from the column in a wide range of concentrations of methanol in chloroform. Indeed, the largest amount is washed off by chloroform-methanol mixture 1:1 vol. (at lower concentrations of methanol mainly by-products are washed off). Note, after column chromatography, chlorine *e*₆ still contains some impurities; the yield of such "crude" chlorine *e*₆ was 29.9 mg (25%).

5. Synthesis of chlorine *e*₆ from 13,15,17-trimethyl ether of chlorine *e*₆.



We dissolved 13,15,17-Trimethyl ether of chlorine *e*₆ (192.0 mg, 0.3 mmol) in 15 ml of tetrahydrofuran and potassium hydroxide (5.04 g, 90 mmol) in 15 ml of water. We added the alkali solution to the macrocyclic compound. We boiled the reaction mixture with a reflux condenser for 3 h, diluted it with water, and neutralised it with dilute hydrochloric acid (7,5 ml HCl_{conc.} in 75 ml water). The precipitate was filtered and dried at room temperature. Column chromatography of the obtained product was performed in silica gel using chloroform-methanol mixtures as eluent. . Indeed, the largest amount chlorine *e*₆ was eluted from the column with a 1:1 vol. chloroform-methanol mixture; the yield of "raw" chlorine was 19.0 mg (10.6%).

Results and discussion

The technique of ester group hydrolysis at position 17 (in the system acetone - hydrochloric acid 10:2 vol., 24 h of stirring at room temperature) was initially used for the synthesis of 17-monocarboxy derivatives of chlorine *e*₆ 13-amides of 15-monomethyl ethers from the corresponding 13-amides of 15,17-dimethyl ethers of chlorine *e*₆ [28]. A modified version of this technique is also well-known - acetone - hydrochloric acid ratio 3:2 vol., stirring time - 3 h (at room temperature) [29]. According to ¹H-NMR, both variants give 17-monocarboxy derivatives as the main product without affecting the 15-complex ester group



and exocycle (in the case of phorbines). However, increasing the time of the modified method up to 4 h causes partial opening of the exocycle in the molecule of methylpheophorbide *a*, despite the main product of the reaction remains pheophorbide *a*. Under similar conditions, pheophorbide *a* (acetone - hydrochloric acid 1:1 vol., 5 h stirring at room temperature) also forms chlorine derivatives in trace amounts. However, contrary to theoretical expectations, methylpheophorbide *a* reacts easily, although it is a more electron-saturated compound compared to pheophorbide proper and contains more ester groups.

The possibility of principal opening the phorbinoxocycle even with such a weak nucleophile as water suggests the process is to be more efficient with the use of stronger nucleophiles. Therefore, we attempted to conduct the opening of the exocycle in the molecule of methylpheophorbide *a* using a solution of an alkali, potassium hydroxide, in methanol. The possibility of simultaneous hydrolysis of ester groups was also assumed. However, methylpheophorbide *a* is very poorly soluble in methanol. Therefore, an organic solvent inert with respect to alkali, tetrahydrofuran (THF), was added to the reaction mixture to increase its solubility. The reaction mixture acquired a bright green colour at the transition of methylpheophorbide *a* into solution. It indicated the opening of the exocycle. TLC of the product isolated by extraction showed as the main product a nonpolar fraction of bright green colour. It moves with the solvent front already at minimum concentrations of methanol (0.5-1% vol.), which is used as eluent in its mixtures with chloroform. In case of monocarboxylic derivatives, the polarity of the product should have been higher, which indirectly supported the formation of trimethyl ester of chlorine e_6 . Subsequently, the formation of trimethyl ether of chlorine e_6 was confirmed by mass spectrometry and $^1\text{H-NMR}$ data. The signal of the phorbinoxocycle proton at 6.28 m.d. disappears and the signal of an additional methyl group at 4.28 m.d. appears in the $^1\text{H-NMR}$ spectrum of the product in CDCl_3 . Hydrolysis of ester groups requires more stringent conditions (larger excesses of alkali, presence of water in solution, elevated temperature), but is complicated by side processes (in particular, decarboxylation of formed chlorine e_6).

We expected that the 13,15,17-trimethyl ether of chlorine e_6 would prove to be the most convenient precursor of free chlorine e_6 (in the form of a triacid). This substance contains three very chemically similar ester groups, that are supposed to undergo hydrolysis under almost identical conditions (unlike phorbines, which contain different reaction centres - at least an exocycle and an ester group). However, experiment shows at longer time of hydrolysis process (3 h in comparison with 1.5 h in case of methylpheophorbide *a*) trimethyl ether of chlorine e_6 gives less yield of the target product (chlorine e_6) and more by-products in comparison with methylpheophorbide *a*. It is confirmed by mass spectrometry (Fig. 6). In addition, chlorine e_6 even after column chromatography still contains the impurities and requires additional purification.

Acid amides can react with nitrous acid formed in the reaction medium from sodium nitrite and mineral acid (most often hydrochloric one). In this process, free acids are obtained from the acid amides.

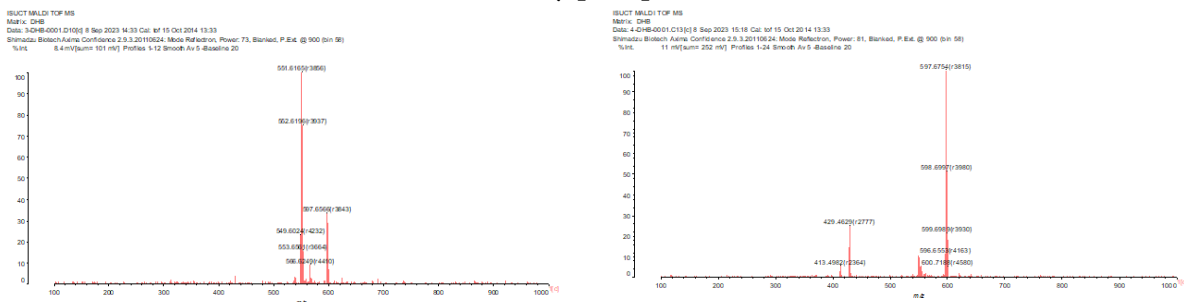
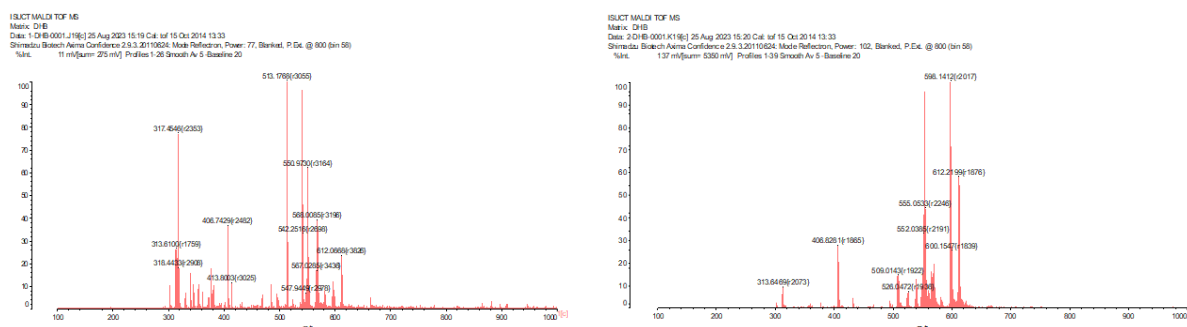
Fraction 1 (CHCl₃ - 20% vol. MeOH)Fraction 2 (CHCl₃ - 50% vol. MeOH)Methylpheophorbide *a*13,15,17-Trimethyl ether of chlorine *e*₆

Fig. 6. Mass spectra of the main fractions obtained during the alkaline hydrolysis of methylpheophorbide *a* and 13,15,17-trimethyl ether of chlorine *e*₆ with potassium hydroxide in the "tetrahydrofuran" system

Hypothetically, chlorine triamide *e*₆ under such conditions should form free chlorine *e*₆, and 13-monoamide 15,17-dimethyl ether of chlorine *e*₆ - the corresponding 13-monocarboxy derivative. However, when we try to use chlorine *e*₆ amides for these purposes, the formation of the corresponding carboxy derivatives is not observed. Moreover, electrophilic substitution in the macrocycle occurs to form a mixture of different 20-substituted derivatives. Product with molecular weight corresponding to isomeric NO²- or ONO-derivatives predominates in the mixture when using excess NaNO₂; while using a stoichiometric amount of nitrite in the mixture remains a significant amount of the initial unreacted amide, containing impurity products of 20-substitution in the macrocycle. According to ¹H-NMR spectroscopy, the signals of all protons, except 20-H, are obtained split, with major signals correspond to the initial amide of chlorine *e*₆. Since the products of 20-substitution do not contain a proton in the indicated position of the macrocycle, they do not give a signal in the ¹H-NMR spectrum.

Conclusions

We have demonstrated phorbixinocycle opening not only by strong O- and N-nucleophiles, but also by such relatively weak O-nucleophiles as alcohols and water. We have studied the conditions for the formation of chlorine *e*₆ and its methyl esters upon treatment of phorbines with potassium hydroxide under various conditions. We have considered possible alternative methods for the preparation and purification of free chlorine *e*₆.



References

1. **Mironov, A.F.** (2004) Current state of chemistry of photosensitisers based on natural porphyrins, chlorines and bacteriochlorines, *Uspexi khimii porfirinov*, 4, pp. 271-289 (in Russian).
2. **Agostinis, P., Berg, K., Cengel, K.A. et al.** (2011) Photodynamic therapy of cancer: an update, *CA Cancer J. Clin.*, 61(4), pp. 250-281. DOI: 10.3322/caac.20114.
3. **Cabuy, E.** (2012) Photodynamic therapy in cancer treatment. RCT summary for professionals, *Reliable cancer therapies. Energy-based therapies*, 3(2), pp. 1-54.
4. **Tsyb, A.F., Kaplan, M.A., Romanko, Y.S. & Popuchiev, V.V.** (2009) *Clinical aspects of photodynamic therapy*. Kaluga: Izd-vo nauch. lit-ry (in Russian).
5. **Van Straten, D., Mashayekhi, V., De Bruijn, H.S., Oliveira, S. & Robinson, D.J.** (2017) Oncologic photodynamic therapy: basic principles, current clinical status and future directions, *Cancers*, 9(19), pp. 1-54. DOI: 10.3390/cancers9020019.
6. **Bonnett, R.** (2000) *Chemical aspects of photodynamic therapy*. Amsterdam: Gordon and Breach Science Publishers.
7. **Kustov, A.V., Berezin, D.B., Strelnikov, A.I. & Lapochkina, N.P.** (2020) *Antitumour and antimicrobial photodynamic therapy: mechanisms, targets, clinical and laboratory studies*. Moscow: Largo (in Russian).
8. **Koifman, O.I. et al.** (2022) Synthetic strategy of tetrapyrrolic photosensitizers for their practical application in photodynamic therapy, *Macroheterocycles*, 15(4), pp. 207-304. DOI: 10.6060/mhs224870k.
9. **Berezin, D.B., Karimov, D.R. & Kustov, A.V.** (2018) *Corroles and their derivatives: synthesis, properties, prospects of practical application*. M.: LENAND (in Russian).
10. **Karimov, D.R., Berezin, D.B., Tomilova, I.K.** (2020) Corroles as aromatic analogs of corrinoids and vitamin B₁₂: synthesis, structural features and macrocycle properties, perspectives of material chemistry on corrole basis, *From Chemistry Towards Technology Step-by-Step*, 1(1), pp. 9-55. DOI: 10.52957/27821900_202020_01_9 [online]. Available at: <http://chemintech.ru/index.php/tor/2020tom1n1> (accessed 10.09.2023).
11. **Pandey, S.K., Sajjad, M., Chen, Y., Zheng, X., Yao, R., Missert, J.R., Batt, C., Nabi, H.A., Oseroff, A.R. & Pandey, R.K.** (2009) Comparative Positron-Emission Tomography (PET) Imaging and Phototherapeutic Potential of ¹²⁴I- Labeled Methyl-3-(1'-iodobenzoyloxyethyl)pyropheophorbide-*a* vs the Corresponding Glucose and Galactose Conjugates, *J. Med. Chem.*, 52, pp. 445-455. DOI: 10.1021/jm8012213.
12. **Zhang, M., Zhang, Z., Blessington, D., Li, H., Busch, T.M., Madrak, V., Miles, J., Chance, B., Glickson, J.D., Zheng, G.** (2003) Pyropheophorbide 2-Deoxyglucosamide: A New Photosensitizer Targeting Glucose Transporters, *Bioconjugate Chem.*, 14, pp. 709-714. DOI: 10.1021/bc034038n.
13. **Varchi, G., Rapozzi, V., Ragno, D., Castoria, G., Di Donato, M., Pietra, E.D., Benfenati, V., Ferroni, C., Guerrini, A., Cesselli, D. & Saracino, E.** (2015) Androgen receptor targeted conjugate for bimodal photodynamic therapy of prostate cancer in vitro, *Bioconjugate Chem.*, 26(8), pp. 1662-1671. DOI: 10.1021/acs.bioconjchem.5b00261.
14. **Tegos, G.P., Anbe, M., Yang, Ch., Demidova, T.N., Satti, M., Mroz, P., Janjua, S., Gad, F. & Hamblin, M.R.** (2006) Protease-Stable Polycationic Photosensitizer Conjugates between Polyethyleneimine and Chlorin(e6) for Broad-Spectrum Antimicrobial Photoinactivation, *Antimicrobial Agents and Chemotherapy*, 50(4), pp. 1402-1410. DOI: 10.1128/aac.50.4.1402-1410.2006.
15. **Hamblin, M.R.** (2016) Antimicrobial photodynamic inactivation: a bright new technique to kill resistant microbes, *Current opinion in Microbiology*, 33, pp. 67-73. DOI: 10.1016/j.mib.2016.06.008.
16. **Kustov, A.V., Morshnev, Ph. K., Kukushkina, N.V., Smirnova, N.L., Berezin, D.B., Karimov, D.R., Shukhto, O.V., Kustova, T.V., Belykh, D.V., Mal'shakova, M.V., Zorin, V.P. & Zorina, T.E.** (2022) Solvation, Cancer Cell Photoinactivation and the Interaction of Chlorin Photosensitizers with a Potential Passive Carrier Non-Ionic Surfactant Tween 80, *Int. J. Mol. Sci.*, 23, pp. 5294-5305 [online]. Available at: <https://doi.org/10.3390/ijms23105294> (accessed 09.10.2023).
17. **Hynninen, P.H.** (1973) Chlorophylls. IV. Preparation and purification of some derivatives of chlorophylls *a* and *b*, *Acta Chem. Scand.*, 27, pp. 1771-1780.



18. **Belykh, D.V.** (2017) Formation of C-O, C-S, C-N and C-C bonds at the periphery of phytochlorines macrocycle during their chemical modification: basic methods and synthetic applications, *Russian Chemical Journal*, LXI(3), pp. 69-109 (in Russian).
19. **Belykh, D.V., Kopylov, E.A., Gruzdev, I.V. & Kuchin, A.V.** (2010) Opening of the exocycle of methylpheophorbide under the action of amines as a one-step method of introduction of additional fragments on the periphery of the chlorine macrocycle, *Zhurn. org. khimii*, 46(4), pp. 584-592 (in Russian).
20. **Nikolaeva, I.A., Misharin, A.Yu., Ponomarev, G.V., Timofeev, V.P. & Tkachev, Ya.V.** (2010) Chlorin e_6 -cholesterol conjugate and its copper complex. Simple synthesis and entrapping in phospholipid vesicles, *Bioorg. Med. Chem. Lett.*, 20, pp. 2872-2875. DOI: 10.1016/j.bmcl.2010.03.041.
21. **Gushchina, O.I., Larkina, E.A., Nikolskaya, T.A. & Mironov, A.F.** (2015) Synthesis of amide derivatives of chlorin e_6 and investigation of their biological activity, *J. Photochem. Photobiol. B: Biology*, 153, pp. 76-81. DOI: 10.1016/j.jphotobiol.2015.09.007.
22. **Hargus, J.A., Fronczek, F.R., Vicente, M.G.H. & Smith, K.M.** (2007) Mono-(L)-aspartylchlorin- e_6 , *Photochemistry and Photobiology*, 83, pp. 1006-1015. DOI: 10.1111/j.1751-1097.2007.00092.x.
23. **Jinadasa, R.G.W., Hu, X., Vicente, M.G.H. & Smith, K.M.** (2011) Syntheses and cellular investigations of 17^3 -, 15^2 -, and 13^1 -amino acid derivatives of chlorin e_6 , *J. Med. Chem.*, 54, pp. 7464-7476. DOI: 10.1021/jm2005139.
24. **Askarov, K.A., Berezin, B.D., Evstigneeva, R.P. et al.** (1985) *Porphyrins: structure, properties, synthesis*. Moscow: Nauka (in Russian).
25. **Taima, H., Okubo, A., Yoshioka, N. & Inoue, H.** (2005) Synthesis of cationic water-soluble esters of chlorin e_6 , *Tetrahedron Lett.*, 46, pp. 4161-4164. DOI: 10.1016/j.tetlet.2005.04.069.
26. **Koifman, O.I. & Ponomarev, G.V.** (2013) *Method for obtaining methylpheophorbide (A)*. 2490273 RF.
27. **Makarov, V.V.** (2023) *Synthesis and physicochemical characteristics of tetrapyrrole macrocycles with polar groups for antimicrobial photodynamic therapy*. PhD. Ivanovo (in Russian).
28. **Tulaeva, L.A., Belykh, D.V., Yakovleva, N.M., Sel'kova, I.A., Rocheva, A.V. & Kuchin, A.V.** (2006) Synthesis and study of chlorophyll derivatives containing a free carboxyl group, *Khimiya i khimicheskaya tekhnologiya*, 49(4), pp. 82-87 (in Russian).
29. **Berezin, M.B.** (1993) *Solvation of chlorophyll and related compounds*. PhD. Ivanovo (in Russian).

Received 27.10.2023

Approved after reviewing 15.11.2023

Accepted 21.11.2023